

Expansion of Bisindole Biosynthetic Pathways by Combinatorial Construction

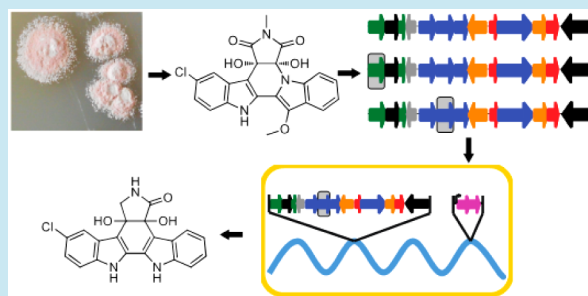
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S Supporting Information

ABSTRACT: Cladoniamides are indolotryptoline natural products that derive from indolocarbazole precursors. Here, we present a microbial platform to artificially redirect the cladoniamide pathway to generate unnatural bisindoles for drug discovery. Specifically, we target glycosyltransferase, halogenase, and oxidoreductase genes from the phylogenetically related indolocarbazole rebeccamycin and staurosporine pathways. We generate a series of novel compounds, reveal details about the substrate specificities of a number of enzymes, and set the stage for future efforts to develop new catalysts and compounds by engineering of bisindole genes. The strategy for structural diversification we use here could furthermore be applied to other natural product families with known biosynthetic genes.

KEYWORDS: *combinatorial engineering, bisindole biosynthesis, glycosyltransferase, halogenase, flavin-dependent monooxygenase, synthetic pathways*



Natural products are an important source of available chemotherapeutics.¹ However, development of new natural product-based drugs has been on the decline over the last decades, for reasons ranging from decreased investment to the structural complexity of natural products and high rates of rediscovery.² With the progress in understanding the biosynthetic pathways of these small molecules and the advances in genetics and genomics, new techniques, such as genome and metagenome mining, have been developed to search new natural products from both culturable and unculturable microbes.^{3,4} Furthermore, approaches such as mutational and combinatorial biosynthesis have been applied to explore new chemical space to prevent the lapse in the discovery pipelines.⁵

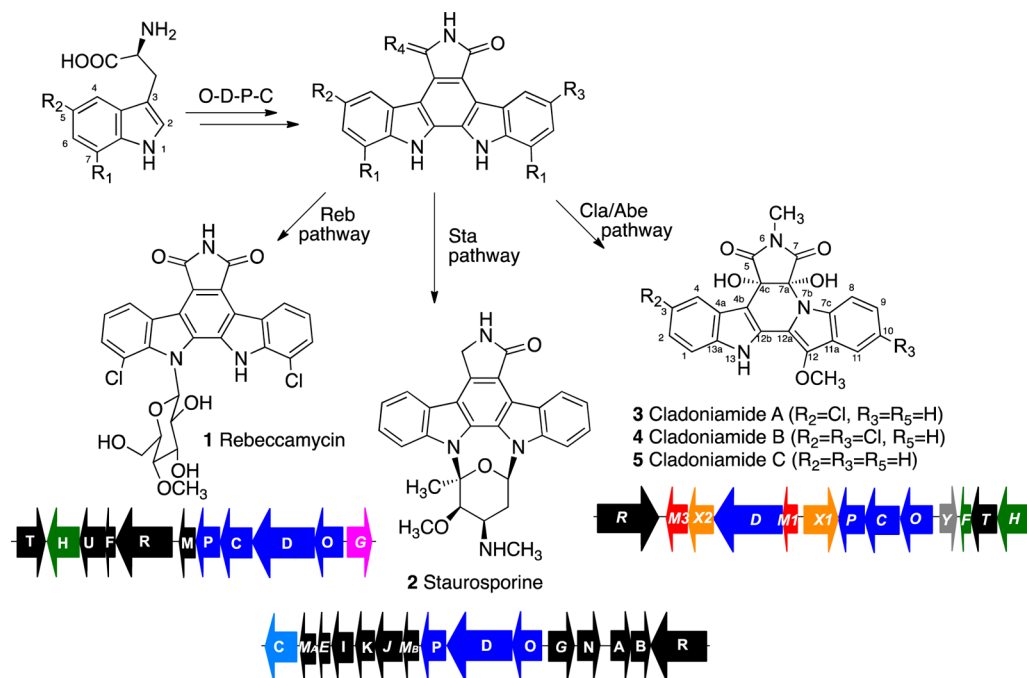
Among microbial natural products, bisindoles that derive from the oxidative dimerization of tryptophan possess diversified biological activities. Well-known examples of such bisindoles include rebeccamycin (**1**), a DNA-topoisomerase I inhibitor, and staurosporine (**2**), a poly-kinase inhibitor, analogues of which progressed into Phase II/III clinical trials against various cancers. Biosynthetic work has established a core set of enzymes, including a chromopyrrolic acid synthase (StaD/RebD), as critical for formation of the indolocarbazole scaffold embedded in molecules such as staurosporine and rebeccamycin.⁶ Recently, by exploiting degenerate primers specific for chromopyrrolic synthase genes, the Brady, Zhang, and Zhu groups isolated a number of new bisindole genes. These efforts have led to the discovery of new indolocarbazoles, such as erdasporines⁷ and streptocarbazoles,⁸ along with nonindolocarbazoles such as spiroindimicins⁹ and methyarcyriarubin,¹⁰ along with the indolotryptolines BE-54017¹¹ and borregomycin A.¹² Structurally, BE-54017 and borregomycin A are closely related to the

indolotryptoline cladoniamides (**3–9**), which were reported by the Andersen and Davies laboratories from a *Streptomyces* strain and proposed to derive from indolocarbazoles.¹³

Indolocarbazoles themselves derive from a four-enzyme pathway from L-tryptophan. This pathway involves (1) production of an indolepyruvate (IPA) imine from L-tryptophan (or a chlorinated derivative) by an amino acid oxidase (StaO/RebO);¹⁴ (2) radical coupling of the IPA enamine by chromopyrrolic acid synthase (StaD/RebD) to give an indolepyruvate enamine dimer, that spontaneously converts to chromopyrrolic acid in oxygen;¹⁵ (3) a cytochrome P450 (StaP/RebP) catalyzed aryl–aryl coupling, giving an unstable intermediate;¹⁶ and (4) interception of a StaP-derived product by a flavin-dependent enzyme (StaC/RebC),¹⁷ ultimately generating the desired indolocarbazole aglycone (Scheme 1, Supporting Information Figure S1). In the case of rebeccamycin and staurosporine, the indolocarbazole scaffolds are then tailored through additional chemistry, including glycosyltransfer and methyltransfer reactions.⁶ A combination of genetic, biochemical, and heterologous biotransformation studies have now revealed that indolotryptolines also derive, as hypothesized,¹³ from the enzymatic processing of indolocarbazole precursors.^{11,18} Indeed, we recently reported the full series of chemical transformations that occur in cladoniamide biosynthesis to convert an indolocarbazole to an indolotryptoline.¹⁸ These transformations consist of (5) *cis*-diol formation across the C4c–C7a double bond by ClaX1; (6) *N*-methylation of the succinimide ring by ClaM1; (7) formation of the indolotryptoline

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Scheme 1. Biosynthesis of Bisindoles^a

^aTwo molecules of L-tryptophan are converted to an indolocarbazole by the action of four enzymes (RebODPC/StaODPC/ClaODPC). Pathway-specific enzymes then generate the final rebeccamycin (1), staurosporine (2), and cladoniamide (3–5) structures. All gene clusters are shown with the *O-D-P-C* genes in blue. Genes investigated in the combinatorial engineering study are chlorinase gene *rebH* (in green), glucosyltransferase gene *rebG* (in pink), and oxidoreductase gene *staC* (light blue). The unique cladoniamide genes are shown in red (methyltransferase genes), orange (flavoenzyme genes *claX1* and *claX2*), green (halogenase and associated reductase gene), gray (unknown, *claY*), and black (regulator and transporter genes). For the rebeccamycin pathway: $R_1=Cl, R_2=R_3=H, R_4=O$; for the staurosporine pathway: $R_1=R_2=R_3=H, R_4=2H$; for the cladoniamide pathway: $R_1=R_5=H, R_2=R_3=H/Cl, R_4=O$.

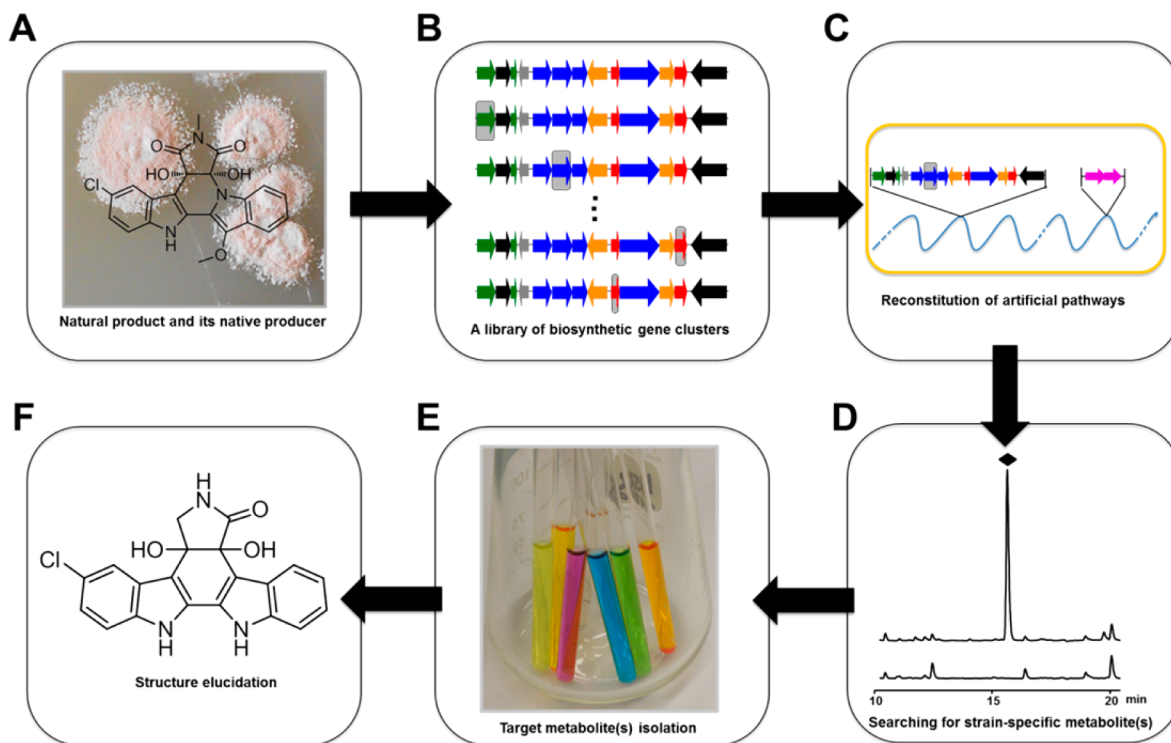
line scaffold by ClaX2; and (8) installation of *O*-methylation by ClaM3 (Supporting Information Figure S1).

Previous studies on the indolocarbazole 1 and 2 gene clusters not only revealed their biosynthetic pathways but also provided a genetic toolbox for combinatorial biosynthesis for production of indolocarbazole compounds.¹⁹ The recent isolation and characterization of a number of new bisindole gene cluster potentially enriches this molecular toolbox. With the biosynthetic gene cluster for cladoniamides (*cla* cluster) in hand, we sought to utilize indolocarbazole genes to artificially redirect the cladoniamide biosynthetic route for synthesis of novel bisindole compounds and to test the substrate specificity of Cla enzymes. The microbial platform we designed for this purpose consists of (i) a “base” biosynthetic gene cluster (*cla* cluster), (ii) selected genes taken from phylogenetic-related clusters, (iii) genetically-, genomically-, and metabolically friendly heterologous hosts (Scheme 2).

First, we generated a small library of engineered *cla* clusters, each of which carries an inactive *cla* gene constructed by targeted gene-inactivation. Introduction of these clusters into heterologous hosts is expected to generate biosynthetic intermediates or shunt products from the cladoniamide pathway. Indeed, our recent studies have shown that introduction of *cla* ($\Delta claM1$) or *cla* ($\Delta claM3$) clusters into *S. albus* or *S. coelicolor* resulted in the detection of a series of bisindole compounds.^{18,20} Next, we reconstituted artificial bisindole pathways by reintroducing intact or engineered *cla* clusters and genes from indolocarbazole gene clusters into *S. albus* or *S. coelicolor*. Specifically, we chose three genes from indolocarbazole systems to introduce into the cladoniamide production system. First, we are interested in the

possibility of generating glycosylated cladoniamides, and we reasoned that the glucosyltransferase RebG, from the rebeccamycin pathway, is the characterized enzyme most likely to accept an indolotryptoline as a substrate. Prior studies had shown some substrate flexibility for this enzyme, including the ability to glycosylate a minimal carbazole ring, as well as a brominated indolocarbazole.²¹ We sought to probe whether RebG was able to glycosylate cladoniamide, or any of the intermediates along the ClaX1–ClaM1–ClaX2–ClaM3 cascade. Second, we targeted RebH, the 7-L-tryptophan halogenase from the rebeccamycin pathway.²² We reasoned that if we eliminated the putative 5-L-tryptophan halogenase gene *claH* from the cladoniamide cluster, introduction of *rebH* might give access to cladoniamides with altered halogenation patterns. Third, we targeted StaC, the flavin-dependent monooxygenase from the staurosporine pathway. This enzyme controls the oxidative outcome in the last step of staurosporine aglycone formation, giving a pyrrolinone ring, in contrast to the maleimide ring observed in rebeccamycin and cladoniamide.^{23,24} Through elimination of *claC* and introduction of *staC*, we anticipated that we could generate cladoniamides and/or cladoniamide intermediates with a pyrrolinone ring.

We first introduced *rebG* into the *S. albus* strain carrying the complete *cla* cluster. High pressure liquid chromatography (HPLC) analysis of the ethyl acetate extract of *S. albus* + *cla* + *rebG* showed four major peaks (Figure 1), two (t_R : 22.0 and 24.1 min) of which were identified to be cladoniamide A (3) and B (4) by comparison with controls (*S. albus* + empty vector and *S. albus* + *cla*, the latter accumulates cladoniamides A, B, C, and D (3–6)²⁰). Another two earlier-eluted peaks (t_R : 17.1 and 19.5 min) displayed UV–vis spectra characteristic of the indolocar-

Scheme 2. Microbial Platform for Combinatorial Construction of New Bisindole Compounds^a

^a(A) Natural product and its native producer. (B) A library of biosynthetic clusters arise from isolation and modification of the natural biosynthetic gene cluster. Gray boxes indicate inactivated genes. (C) Reconstitution of the artificial pathways in optimized heterologous hosts. (D) Metabolic analysis for strain-specific metabolite(s). (E) Large-scale fermentation for target compound(s) purification. (F) Structure elucidation.

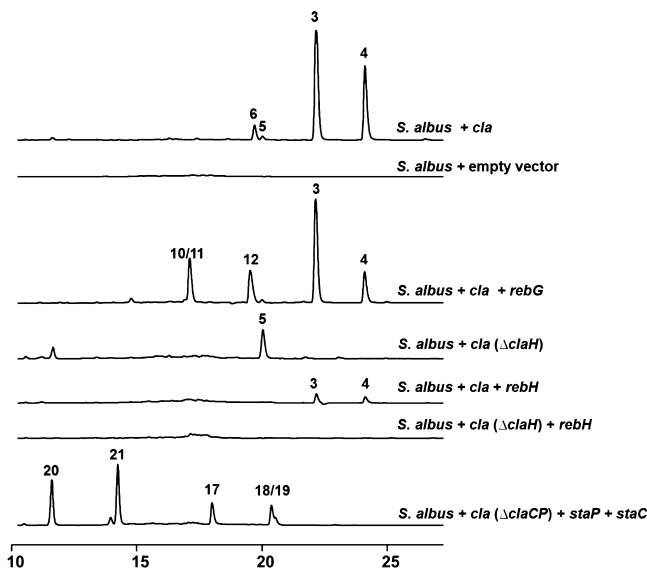


Figure 1. Analytical HPLC profiles of various genetic mutants constructed for the combinatorial study (detection wavelength = 348 nm).

bazole chromophore and were characterized as a mixture of glucosyl-3-chloroarcyriaflavin A (10) and glucosyl-9-chloroarcyriaflavin A (11) and pure glucosyl-3,9-chloroarcyriaflavin A (12) by MS and NMR analysis (Supporting Information Tables S3–S5 and Figures S2–S3). These three molecules are new analogues of demethylrebecamycin. These results suggest that RebG favors indolocarbazole substrates and cannot readily

glycosylate either cladoniamide A or B, but RebG is able to accept alternately chlorinated indolocarbazoles.

Next, to probe whether we could modify the natural *cla* pathway to obtain new bisindoles with chlorine atom(s) at alternative positions, the *cla* ($\Delta claH$) cluster was transferred into both *S. albus* and *S. coelicolor* strains. From the *S. albus* + *cla* ($\Delta claH$) heterologous production system, cladoniamide C (5) was identified to be the major product (Figure 1), while from the *S. coelicolor* + *cla* ($\Delta claH$) system, along with the production of cladoniamide C (5), two new metabolites (13 and 14) were also produced (Figure 2). They were identified as new, nonchloro derivatives of cladoniamides D/E and F/G by UV/vis and LC-MS analysis (Figure 2; Supporting Information Table S3 and Figure S4). Next, *rebH* was introduced into *S. albus* + *cla* ($\Delta claH$), but no cladoniamides or new products were detected. We also coexpressed *rebH* with the intact *cla* cluster in *S. albus*, but this strain produced a very low amount of cladoniamides A and B (Figure 1). These results suggest that early Cla enzymes might not be able to accept 7-chloro-L-tryptophan, and the addition of RebH could hijack L-tryptophan from the *cla* pathway.

Finally, StaC has been shown to be a branch point where it diverges the *sta* pathway from *reb/cla* pathways. We sought to probe the possibility of artificially redirecting the product of StaC (K252c, 17) back to *cla* pathway to generate bisindoles with staurosporine-like pyrrolinone rings. To achieve this, we first constructed an *S. albus* strain harboring the *cla* ($\Delta claC$) cluster, in which *claC* was in-frame deleted. This strain unexpectedly accumulated 9-chloro-chromopyrrolic acid (15) and 9,9'-dichloro-chromopyrrolic acid (16) (Supporting Information Figures S5–S6), instead of producing the chlorinated derivatives of arcyriaflavin A, K252c, and 7-hydroxy-K252c,²⁵ suggesting a

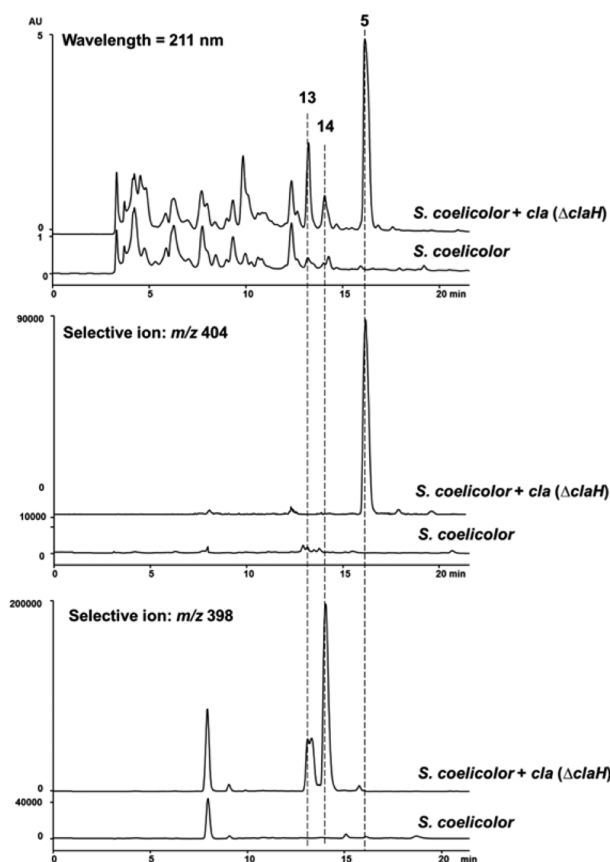


Figure 2. Production of nonchlorinated cladoniamides C (5), H (13), and I (14) from engineered strain *S. coelicolor* YD17. *S. coelicolor* YD17 produces 5 as a major clone-specific metabolite along with cladoniamide H (13) and I (14) (detection wavelength = 211 nm). Selective ion monitoring at $m/z = 404$ reveals 5 as clone-specific metabolite. Selective ion monitoring at $m/z = 398$ reveals 13 and 14 as clone-specific metabolites.

polar effect on the downstream *claP* gene. We thus introduced both *staP* and *staC* to the *S. albus* + *cla* ($\Delta claCP$) system, in order to construct a pathway that would provide staurosporine aglycone (K252c, 17) or its chlorinated derivatives for the downstream Cla enzymes. Extracts of *S. albus* + *cla* ($\Delta claCP$) + *staPC* had five major clone-specific metabolites compared to control strains (Figure 1). The compound 17 (t_R : 18.0 min) showed identical UV-vis spectrum as K252c, and it was characterized as K252c by comparing its LC-MS and 1H NMR data (Supporting Information Table S3 and Figure S7) with the reported data from K252c.²⁶ The HPLC peaks for compounds 18 and 19 are largely overlapped, and both of them display similar UV-vis spectra as K252c (17) (Supporting Information Figure S4). They were collected together by semiprep HPLC and analyzed by ESI-MS, revealing a single MS signal (m/z 346, $[M + H]^+$) with the signature of a chlorine atom. They were identified to be 3-chloro and 9-chloro derivatives of K252c based on the 1H NMR data (Supporting Information Figure S7). Compounds 20 (t_R : 11.6 min) and 21 (t_R : 14.2 min) were identified as 4c/7a diol derivatives of K252c and Cl-K252c based on their MS, 1D and 2D NMR data (Supporting Information Tables S6–S7 and Figures S8–S9). Interestingly, in contrast to Cl-K252c, which we isolated as a mixture of 3-chloro and 9-chloro K252c (18/19), the 4c/7a diol derivative of Cl-K252c (21) was purified as single component. The structure of 21 implies that ClaX1 can accept only one of the two Cl-K252c molecules as a substrate, and the

accumulation of 21 furthermore suggests that the downstream enzyme ClaM1 cannot utilize compound 21 as a substrate (Supporting Information Figure S1). To further solidify the above conclusions, *E. coli*-based biotransformation assays were performed. Specifically, the mixture of 3-chloro and 9-chloro derivatives of K252c (18/19) was fed to the culture of *E. coli* strain expressing ClaX1, and metabolic profiling by HPLC revealed the production of 21. Furthermore, only one of the two Cl-K252c's was consumed (Figure 3). Similarly, 20 was produced when K252c (17) was fed to this strain. Both 20 and 21 are new compounds.

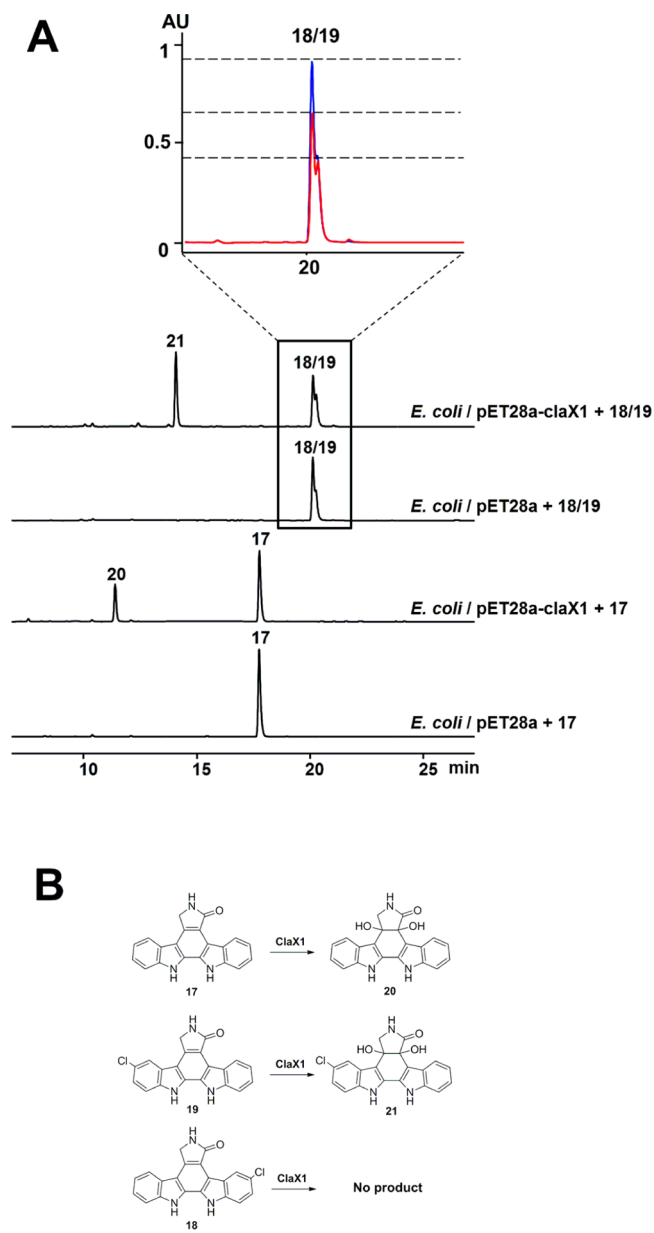
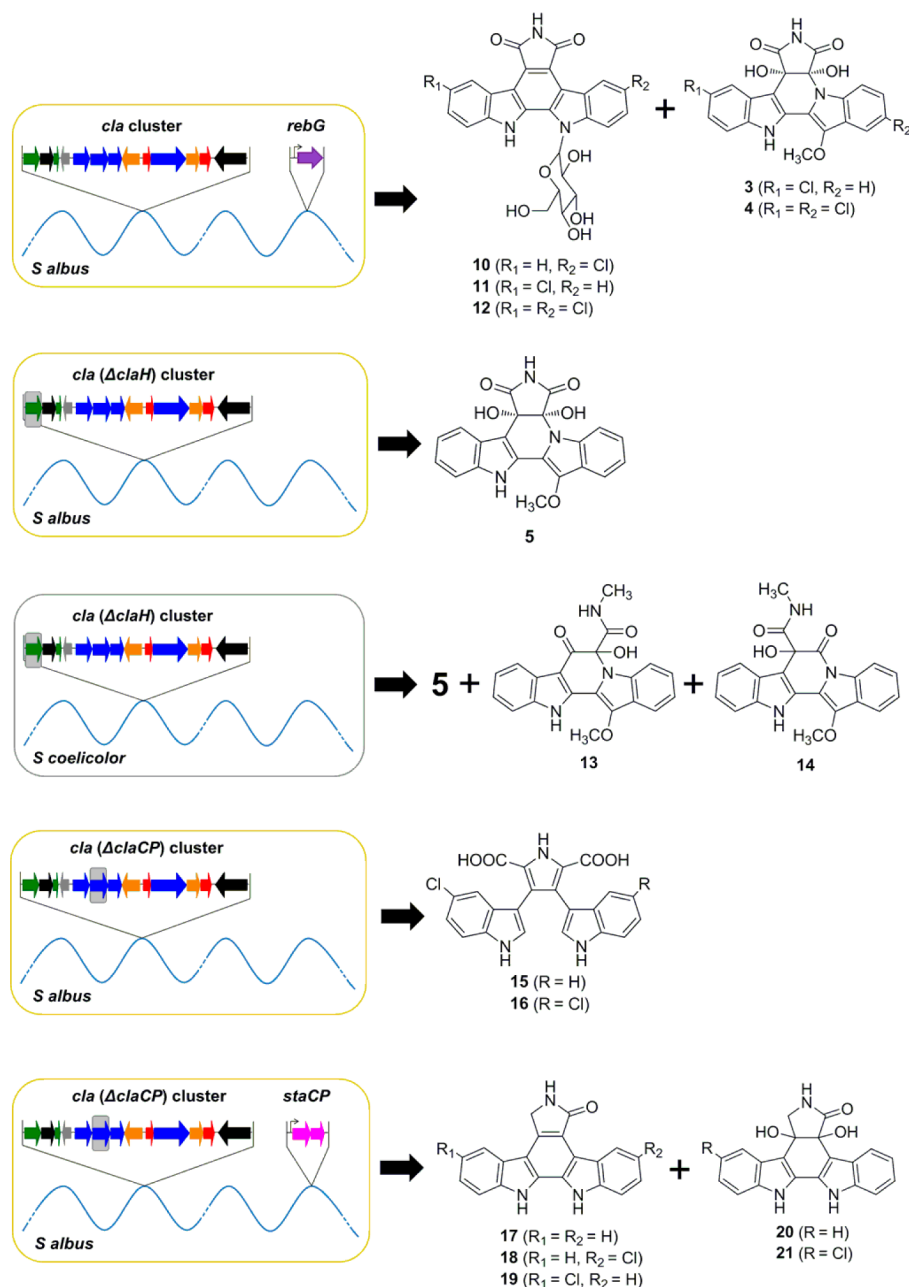


Figure 3. *In vivo* biotransformation of 17 and 18/19 with *claX1*. (A) HPLC traces for *in vivo* biotransformation assay. Insert, only the peak at the left was consumed in the *in vivo* biotransformation assay of *E. coli*/pET28a-*claX1* + 18/19, which is assumed to be 19, based on the corresponding production of 21. Blue, *E. coli*/pET28a + 18/19. Red, *E. coli*/pET28a-*claX1* + 18/19. Detection wavelength: 348 nm. (B) Scheme for conversion by ClaX1.

Scheme 3. Combinatorial Biosynthesis of Bisindole Metabolites



We tested the biological activities of selected new compounds generated from our combinatorial platform, and we observed IC_{50} values of $1.42 \mu M$ for 10/11, $3.24 \mu M$ for 12, and $4.42 \mu M$ for 18/19 against colon cancer cell line HCT-116.

Microbial natural products and their biosynthetic gene clusters are shaped by long evolutionary processes, involving joining of subclusters and gene mutation, acquisition, loss, duplication, and functional divergence.^{27,28} Understanding the biosynthetic pathways of natural products that putatively share the same ancestor can give insights into the naturally occurring modes of structural diversification, and facilitate efficient manipulation of these pathways for the generation of unnatural products. Previous studies by Brady and our group established that the rare indolotryptoline scaffolds derive from more common indolocarbazole structures. In this study, we introduce *rebG*, the glycosyltransferase from the rebecamycin pathway into a

cla-containing strain. Our results show that RebG can intercept the chlorinated arcylriaflavin A intermediates from the cladoniamide pathway to generate new glycosylated indolocarbazoles (Scheme 3). However, the enzyme appears unable to react with cladoniamide itself, or any intermediate beyond the ClaX1 substrate, likely reflecting its need for a substrate with an intact carbazole.²¹ In a next set of experiments, we show that elimination of the chlorinase gene *claH* can give the full suite of nonchlorinated cladoniamides. However, halogenation at the 7 position of *L*-tryptophan, through introduction of 7-*L*-tryptophan chlorinase *rebH*, apparently hijacks tryptophan precursors away from the cladoniamide pathway, suggesting that early cladoniamide enzymes are unable to accept 7-chloro-*L*-tryptophan precursors (Scheme 3). Finally, we interrogate introduction of a pyrrolinone-containing indolocarbazole into the cladoniamide pathway through deletion of *claC* and introduction of *staPC*. We

find that ClaX1 can accept an indolocarbazole with a pyrrolinone ring to generate **20** or **21**; however, downstream enzyme(s) are unable to accept substrates with the pyrrolinone ring. Interestingly, whereas StaP/StaC appear to be able to process a substrate with chlorine at either the 3 or 9 position, ClaX1 is highly specific to the positioning of chlorine on the ring, tolerating chlorination at the 3 position but not the 9 position (Scheme 3). Overall, our results suggest that unique substrate specificities have been molded into cladoniamide and indolocarbazole enzymes during their distinct evolutionary histories, making them best suited to react with native substrates.

The platform developed for this study gave access to a number of new compounds, including glucosyl-3,9-dichloroarxyriaflavin A (**12**) and the 4c/7a diol of Cl-K252c (**21**). Additionally, this platform should be highly suitable for precursor-directed biosynthesis. For instance, feeding of synthetic indolocarbazole precursors to the *S. albus* + *cla* (Δ *claCP*) system should allow us to generate new compounds and enable us to further assess substrate tolerance of the Cla enzymes late in the pathway. Finally, this combinatorial work contributes to knowledge about bisindole enzyme substrate specificities, laying the groundwork for future efforts to engineer bisindole enzymes for higher substrate tolerance, to ultimately enable the production of an even larger suite of bisindole molecules.

METHODS

Combinatorial Engineering.

- (i) For construction of a library of engineered *cla* clusters, vector pYD1 carrying the whole biosynthetic gene cluster was subjected to a series of targeted gene-inactivation protocols by using REDIRECT technique,²⁹ resulting in modified *cla* clusters, including *cla* (Δ *claH*) and *cla* (Δ *claC*).
- (ii) For the construction of strain *Streptomyces* + *cla* cluster + *rebG*, the coding region of *rebG* was amplified from the genomic DNA of *Lechevalieria aerocolonigenes* DSM 44217 using Phusion DNA polymerase. NdeI and XbaI double digested PCR products were inserted into the corresponding sites of pYLD20,³⁰ resulting in pYLDrebG. After confirmation by DNA sequencing, the plasmids were introduced into *Streptomyces* strains carrying *cla* cluster by conjugation.³¹ Thiostrepton-, apramycin-, and kanamycin-resistant clones were confirmed for site-specific integration by PCR with the primer pairs for *rebG* (Supporting Information Table S2).
- (iii) For the construction of strain *Streptomyces* + *cla* (Δ *claH*) cluster, *Streptomyces* + *cla* (Δ *claH*) cluster + *rebH*, and *Streptomyces* + *cla* cluster + *rebH*, the vector carrying the *cla* (Δ *claH*) cluster was passaged through *E. coli* ET12567/pUZ8002 before being introduced via conjugation into *S. albus* J1074³² and *S. coelicolor* M1146³³ to afford *S. albus* YD22 and *S. coelicolor* YD17, respectively. Clones were confirmed for integration by PCR with the primer pairs for *claH* (Supporting Information Table S2). The coexpression of *rebH* with the intact or engineered *cla* cluster was performed similarly as described for the coexpression of *rebG*.
- (iv) For the construction of strains *Streptomyces* + *cla* (Δ *claCP*) cluster and *Streptomyces* + *cla* (Δ *claCP*) cluster + *staPC*, the vector carrying the *cla* (Δ *claC*), in which *claP* was also inactivated due to a polar effect, was introduced into *S. albus* J1074 to generate *S. albus* YD23. The coding regions

of *staP* and *staC* genes were amplified from pET vectors (generously provided by the Walsh and Drennan laboratories) using Phusion DNA polymerase. NdeI and XbaI double digested PCR products of *staC* and XbaI-digested PCR products of *staP* were inserted into the corresponding sites of pYLD20, resulting in pYLDstaCP. After confirmation by DNA sequencing, the plasmid was introduced into *S. albus* YD23 by conjugation. Thiostrepton-, apramycin-, and kanamycin-resistant clones were confirmed for site-specific integration by PCR with the primer pairs for *staP* and *staC* (Supporting Information Table S2).

Production and Analysis of Metabolites from Engineered *Streptomyces* Strains. Cultivation of *Streptomyces* mutants was performed as described previously.²⁰ At the end of fermentation, the cultures were extracted twice with 1 volume of ethyl acetate, and combined extracts were dried under vacuum and then dissolved in methanol.

Analysis of bisindole compounds production was performed with an Agilent 1260 high-pressure liquid chromatography (HPLC) apparatus with a diode array UV detector, fitted with an Phenomenex Luna C18(2) column (5 μ m, 4.6 mm inner diameter (ID) \times 250 mm). Elution was performed at 1 mL/min with a mobile-phase mixture consisting of a linear gradient of water and acetonitrile (85:15 to 20:80, 0 to 25 min; 0:100, 25 to 30 min), both of which contain 0.05% (v/v) trifluoroacetic acid. LC-MS was performed under the same conditions (except that the flow rate was set at 0.8 mL/min) on an Agilent 6120 Quadrupole LC/MS system operated in both positive and negative ion electrospray modes.

Isolation, Purification, and Structural Elucidation of Bisindoles from *Streptomyces* Mutants. Ethyl acetate extracts from *Streptomyces* cultures (3–6 L) were dried under vacuum and then dissolved in methanol. The resulting crude extracts were fractionated on reversed-phase C8 flash column with MeOH/water gradient elution and Sephadex LH-20 with MeOH elution. Metabolites of interest, as tracked by analytical HPLC as described above, were purified from these fractions by reversed-phase semipreparative HPLC (Phenomenex Luna C18(2), 5 μ m, 10 mm ID \times 250 mm) (Supporting Information Figure S10). Compounds **10/11** (yield: 5.5 mg/L) were purified using 30:70 acetonitrile/water. Compound **12** (yield: 5 mg/L) was purified using 37:63 acetonitrile/water. Compound **15** (yield: 6.8 mg/L) was purified using 54:46 acetonitrile/water. Compound **16** (yield: 8 mg/L) was purified using 58:42 acetonitrile/water. Compound **17** (yield: 2.4 mg/L) was purified using 60:40 acetonitrile/water. Compound **18/19** (yield: 2 mg/L) were purified using 70:30 acetonitrile/water. Compound **20** (yield: 4.5 mg/L) was purified using 45:55 acetonitrile/water. Compound **21** (yield: 5 mg/L) was purified using 50:50 acetonitrile/water.

The ¹H- and ¹³C NMR spectra were recorded on a Bruker AV-600 MHz spectrometer with a 5 mm CPTCI cryoprobe. The DMSO-*d*₆ signals (2.50 ppm for ¹H and 39.5 ppm for ¹³C) were used as references. High-resolution mass spectrometry data were acquired using a Micromass LCT time-of-flight (TOF) mass spectrometer equipped with an electrospray ion source. Structure elucidation of **10**, **11**, **12**, **20**, and **21** is described in Supporting Information Methods.

■ ASSOCIATED CONTENT**5 Supporting Information**

Supporting tables, figures, methods, and references. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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Notes

The authors declare no competing financial interest.

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